

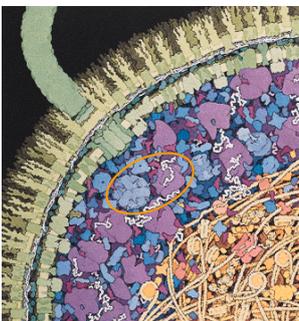
From protein structure prediction to mechanistic insights into signaling and disease diagnostics

Holger Gohlke




Mission

Understand, predict, and modulate molecular and cellular behavior from an atomistic perspective



Taken from David S. Goodsell, *The Machinery of Life*, 1997.

Application studies

$\alpha_5\beta_1$ integrin
Gohlke, Häußinger, Hepatology 2013
Reinehr, Gohlke, Häußinger, JBC 2010

$\alpha_{IIb}\beta_3$ integrin
Donner, Gohlke, Evers, Cell Signal 2018
Pagani, Gohlke, Sci Rep 2016
Pagani, Scharf, Gohlke, JBC 2018
Donner, Gohlke, Elvers, Sci Signal 2016

TGR5
Greife, Gohlke, Kettel, Seidel, Sci Rep 2016
Gertzen, Kettel, Gohlke, Eur J Med Chem 2015
Spomer, Gohlke, Kettel, JBC 2014

Glutamine synthetase
Frieg, Häußinger, 06

MDF
Tanabe, Gohlke, U
Sachs, Gohlke, T
Droge, Gohlke, K

ET
Schott-Verdugo, Gohlke, Groth, Sci Rep 2018

PEPCK
Nguyen, Gohlke, Groth, Sci Rep 2017

PPDK
Cupka, Gohlke, Sci Rep 2017
Minges, Gohlke, Groth, Sci Rep 2017

HDAC
Raudszus, Gohlke, Hansen, BMC 2019
Radlmg, Gohlke, Hansen, Med Chem Comm 2015
Krieger, Gohlke, Hansen, J Med Chem 2017
Stenzel, Gohlke, Kurz, J Med Chem 2017
Diederich, Gohlke, Hansen, Chem Comm 2016
Marek, Gohlke, Kurz, J Med Chem 2013

NHR2
Schanda, Gohlke, Wichmann, Haematol 2017
Metz, Wichmann, Gohlke, JOM 2013
Wichmann, Gohlke, Grez, Blood 2010

Hsp90
Bhatia, Gohlke, Hauer, Blood 2018
Krieger, Gohlke, Hansen, Chem Eur J 2017
Diedrich, Gohlke, Hansen, Chem Eur J 2016
Spanier, Gohlke, Kurz, J Org Chem 2014
Boop, Gohlke, Jose, BSA - Gen Subj 2016
Cigla, Groth, Gohlke, PLOS ONE 2014

Lipases
Frieg, Häußinger, 06
Tanabe, Gohlke, U
Sachs, Gohlke, T
Droge, Gohlke, K

Glutamine synthetase
Frieg, Häußinger, 06

MDR3/Pdr5
Tanabe, Gohlke, Lamping, Antim. A. Chemo. 2019
Sachs, Gohlke, Teusch, Front Pharmacol 2019
Droge, Gohlke, Kettel, J Hepatol 2017

ETR1
Schott-Verdugo, Gohlke, Groth, Sci Rep 2019
Mlic, Gohlke, Groth, Sci Rep 2018

PEPC
Nguyen, Gohlke, Groth, Sci Rep 2016

PPDK
Cupka, Gohlke, Sci Rep 2017
Minges, Gohlke, Groth, Sci Rep 2017

FRET
Zhang, Gohlke, Mink, Retrovirology 2016
Sanabria, Gohlke, Seidel, Nature Comm 2020
Dimura, Gohlke, Seidel, Curr Opin Struct Biol 2018
Kainr, Gohlke, Seidel, Nature Methods 2012



Application studies

$\alpha_5\beta_1$ integrin
Gohlke, Häußinger, Hepatology 2013
Reinehr, Gohlke, Häußinger, JBC 2010

$\alpha_{IIb}\beta_3$ integrin
Donner, Gohlke, Evers, Cell Signal 2018
Pagani, Gohlke, Sci Rep 2016
Pagani, Scharf, Gohlke, JBC 2018
Donner, Gohlke, Elvers, Sci Signal 2016

TGR5
Greife, Gohlke, Kettel, Seidel, Sci Rep 2016
Gertzen, Kettel, Gohlke, Eur J Med Chem 2015
Spomer, Gohlke, Kettel, JBC 2014

Glutamine synthetase
Frieg, Häußinger, Gohlke, PLOS Comp Biol 2016

MDR3/Pdr5
Tanabe, Gohlke, Lamping, Antim. A. Chemo. 2019
Sachs, Gohlke, Teusch, Front Pharmacol 2019
Droge, Gohlke, Kettel, J Hepatol 2017

ETR1
Schott-Verdugo, Gohlke, Groth, Sci Rep 2019
Mlic, Gohlke, Groth, Sci Rep 2018

PEPC
Nguyen, Gohlke, Groth, Sci Rep 2016

PPDK
Cupka, Gohlke, Sci Rep 2017
Minges, Gohlke, Groth, Sci Rep 2017

HDAC
Raudszus, Gohlke, Hansen, BMC 2019
Radlmg, Gohlke, Hansen, Med Chem Comm 2015
Krieger, Gohlke, Hansen, J Med Chem 2017
Stenzel, Gohlke, Kurz, J Med Chem 2017
Diederich, Gohlke, Hansen, Chem Comm 2016
Marek, Gohlke, Kurz, J Med Chem 2013

NHR2
Schanda, Gohlke, Wichmann, Haematol 2017
Metz, Wichmann, Gohlke, JOM 2013
Wichmann, Gohlke, Grez, Blood 2010

Hsp90
Bhatia, Gohlke, Hauer, Blood 2018
Krieger, Gohlke, Hansen, Chem Eur J 2017
Diedrich, Gohlke, Hansen, Chem Eur J 2016
Spanier, Gohlke, Kurz, J Org Chem 2014
Boop, Gohlke, Jose, BSA - Gen Subj 2016
Cigla, Groth, Gohlke, PLOS ONE 2014

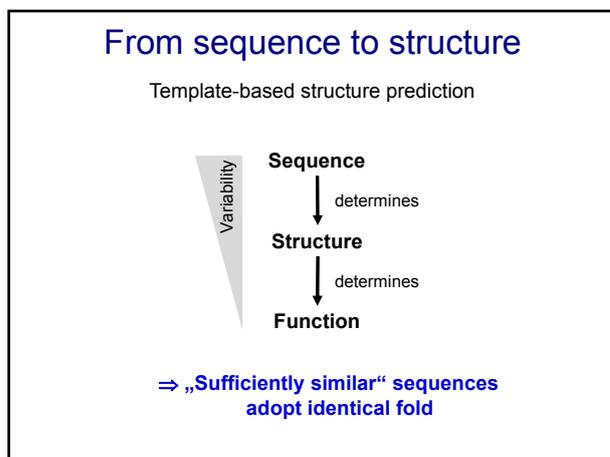
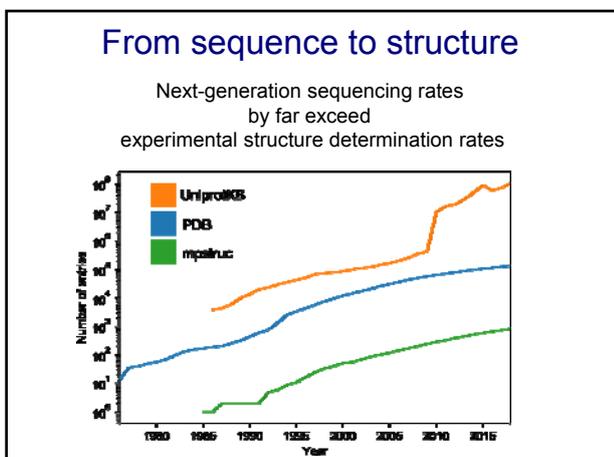
Lipases
Verma, Jaeger, Gohlke, J Comp Chem 2019
Stoczninski, Gohlke, Jaeger, Microb Cell Fact 2017
Rath, Jaeger, Gohlke, PLOS Comp Biol 2016
Rath, Jaeger, Gohlke, PLOS ONE 2015

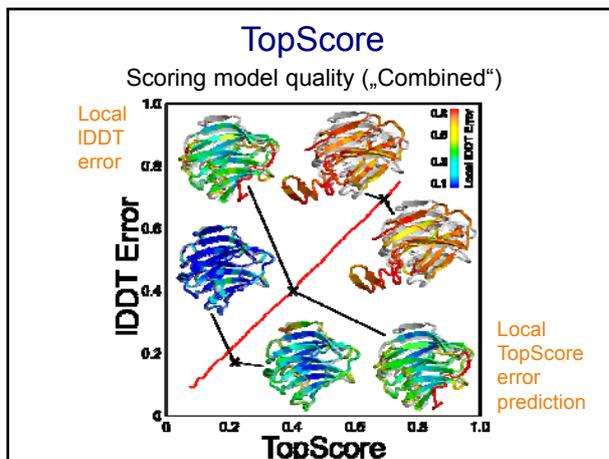
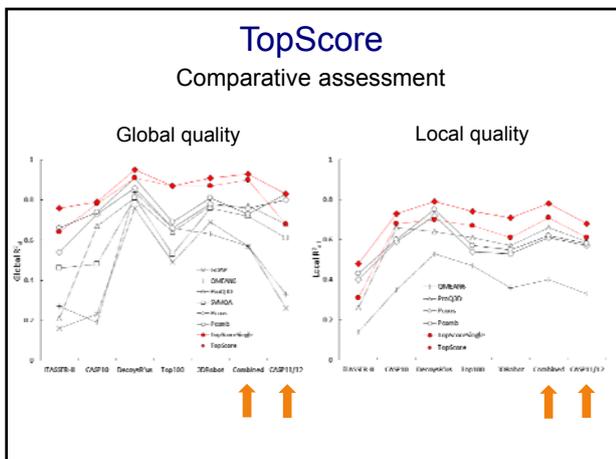
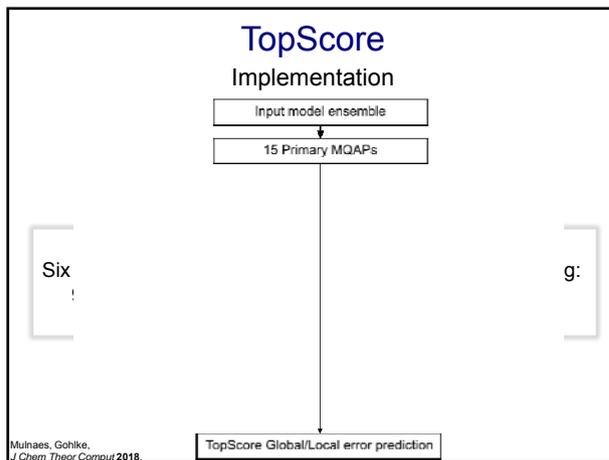
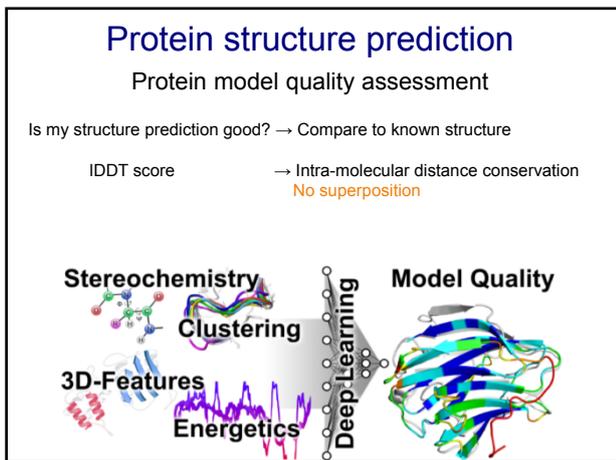
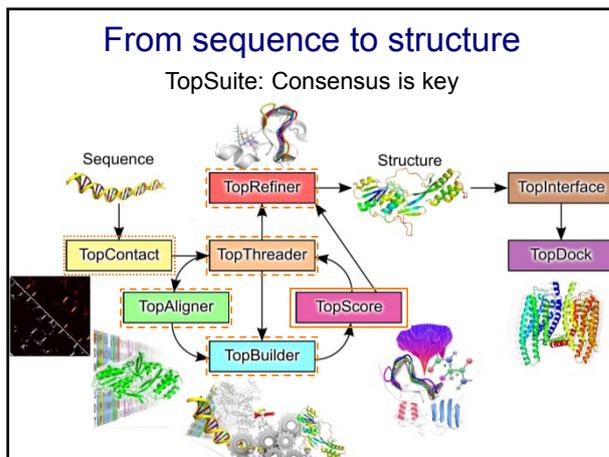
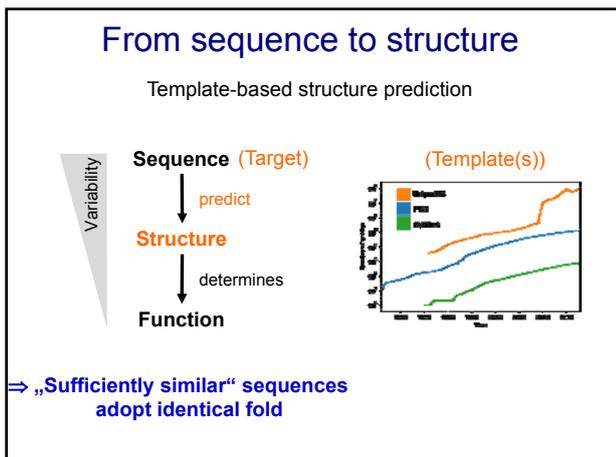
Aldolases
Bisterfeld, Gohlke, Pietruszka, PLOS ONE 2016
Dick, Gohlke, Pietruszka, Sci Rep 2016

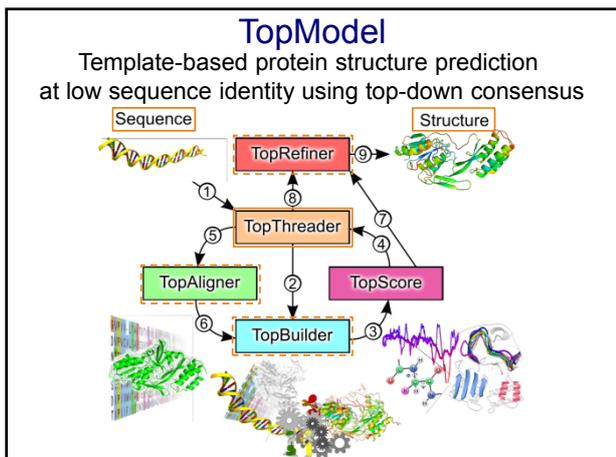
NSR
Porta, Smits, Gohlke, BMC 2019
Khosha, Gohlke, Smits, PLOS ONE 2016
Khosha, Gohlke, Smits, Sci Rep 2016

Vifs
Gu, Gohlke, Mink, J Virol 2018
Zhang, Gohlke, Mink, Retrovirology 2016

FRET
Sanabria, Gohlke, Seidel, Nature Comm 2020
Dimura, Gohlke, Seidel, Curr Opin Struct Biol 2018
Kainr, Gohlke, Seidel, Nature Methods 2012







TopModel

Finding templates

The consensus problem:
Predictor performance is **case-dependent** → No one is always right

Which are the best template(s) for my protein?

- Let **DNNs** find the *best* hit
- Top-down consensus** → Discard hits that do not fit the *best* hit

Predictor	ATM100 < 5 (%)	ATM100 5-15 (%)	ATM100 > 15 (%)
HMMER3	100	0	0
HHBLITS	100	0	0
FASTA36	100	0	0
SAMT2K	100	0	0
DELTA-BLAST	100	0	0
pDom Threader	100	0	0
HHSearch	100	0	0
pGen Threader	100	0	0
RAPTOR-X	100	0	0
LONETS	100	0	0
SPARKS-X	100	0	0
FFAS03	100	0	0
TopThreader	~40	~40	~20

TopModel

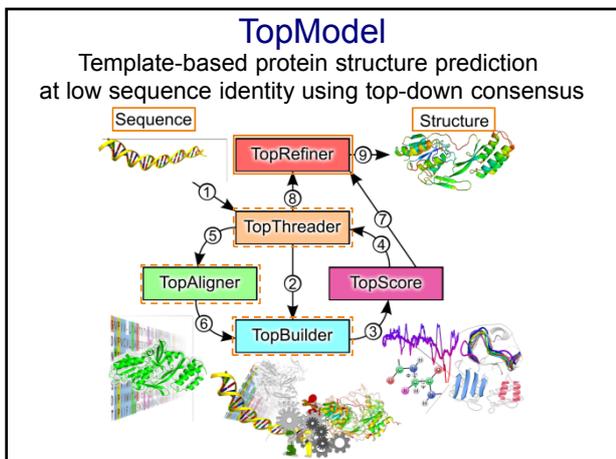
Finding templates

- Primary threading ⇒ Template(s) + score(s)

TopModel

Finding templates

- Primary threading
- Pre-filtering
- Alignment fitting
- Score templates (DNN)
- Redundancy clustering
- False positive removal (DNN)
- Consensus generation
- Ranking (DNN)



TopModel

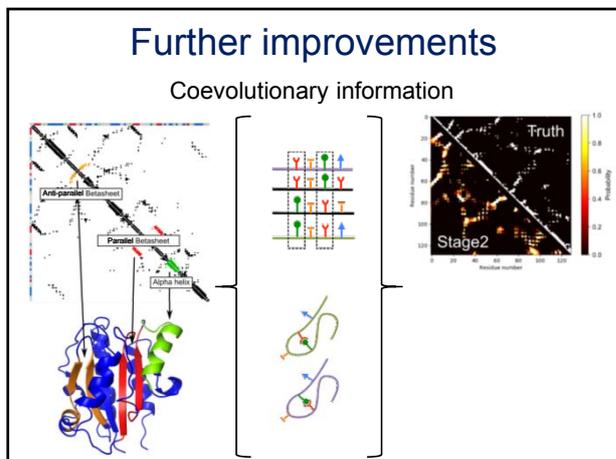
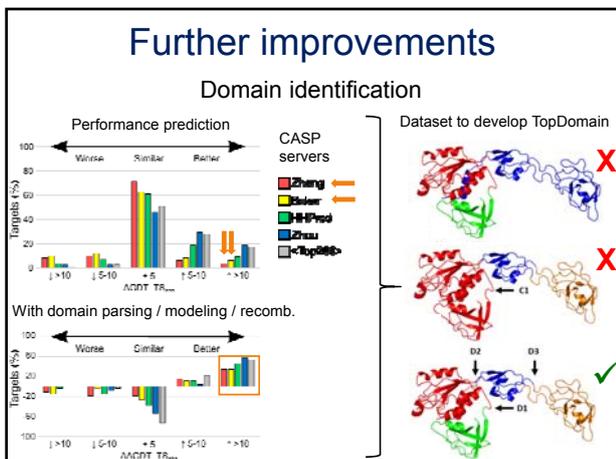
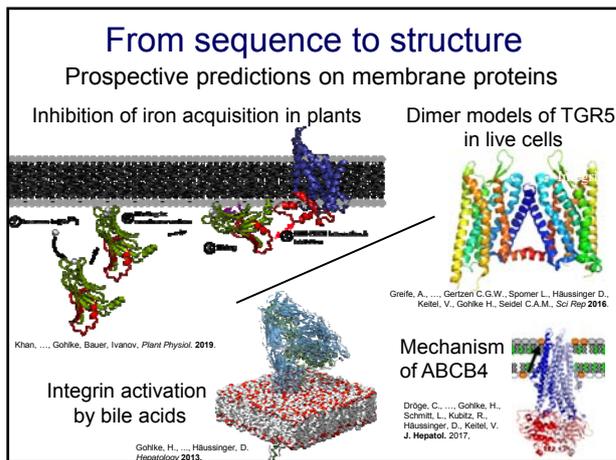
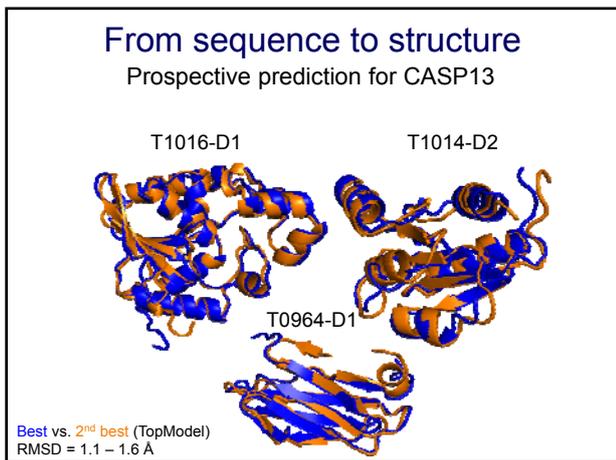
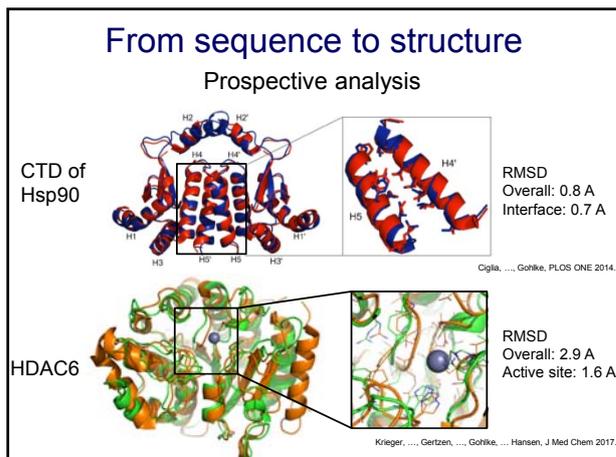
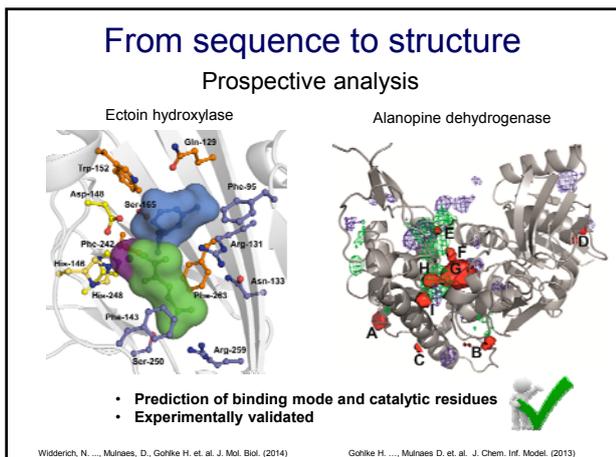
Refining solutions

- TopScore tells us **where** we are wrong
- Delete bad regions** from initial predictions
- Put good pieces together** into new models and repeat

Advantages: Converge on the best - not on the average
Disadvantages: Speed

Prospective modeling of SaNSR

Predictor	GDT_TS (%)
SPARKS-X	0.36
FFAS03	0.32
HHSearch	0.40
RAPTOR-X	0.49
TopModel	0.55



Conclusion



TopScore
JCTC 2018.



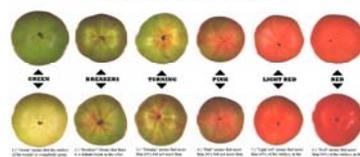
TopModel
JCTC 2020.

Much can (still) be gained in protein structure prediction by

- optimally combining different methods,
- using 'big data' training sets and
- deep neural networks.

Ethylene and ripening

COLOR GUIDE FOR: TOMATOES

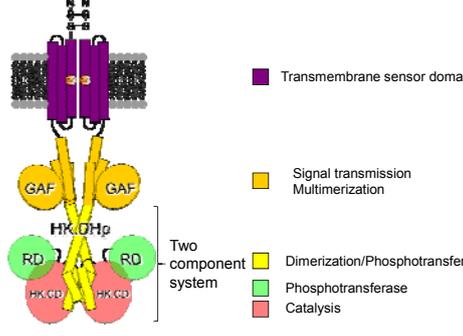


$$\begin{array}{c}
 H & & H \\
 & \backslash & / \\
 & C = C \\
 & / & \backslash \\
 H & & H
 \end{array}$$

- Reducing concentration of ethylene extends 60% postharvest life
- Efforts:
 - Molecular competitors
 - Ethylene absorbers
 - Temperature

<https://youtu.be/k3afpyUw10>

Ethylene ETR1

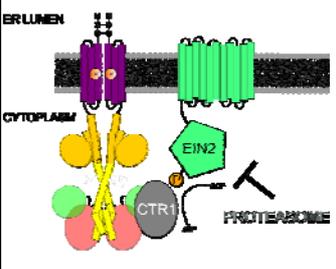


- Transmembrane sensor domain
- Signal transmission
- Multimerization
- Dimerization/Phosphotransferase
- Phosphotransferase
- Catalysis

Two component system

Mayerhofer, H., Panesarvelam, S., Kallonen, H., Tuokkanen, A., Mertens, H. D. T., & Mueller-Dieckmann, J. (2016).
Milo, D., Dick, M., Muhias, D., Pfeifer, C., Kinnari, A., Gohlke, H., & Groth, G. (2018).

ETR1 - Mechanism of action

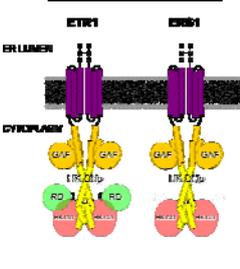


ER LUMEN
 CYTOPLASM
 EIN2
 CTR1
 PROTEASOME

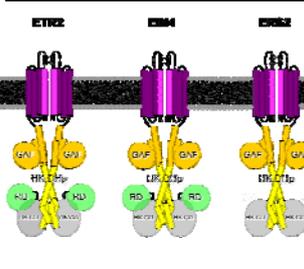
Light, K. M., Wianiewski, J. A., Vinyard, W. A., & Kieber-Emmons, M. T. (2016).

Ethylene receptor families

Subfamily 1

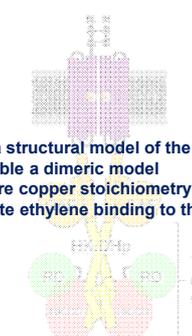


Subfamily 2



Chenail, S. N., Wang, X., Binder, B. M., & Schaller, G. E. (2013).

ETR1 domains

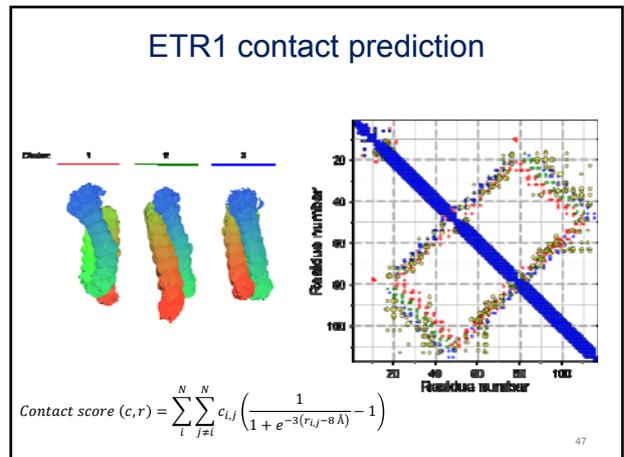
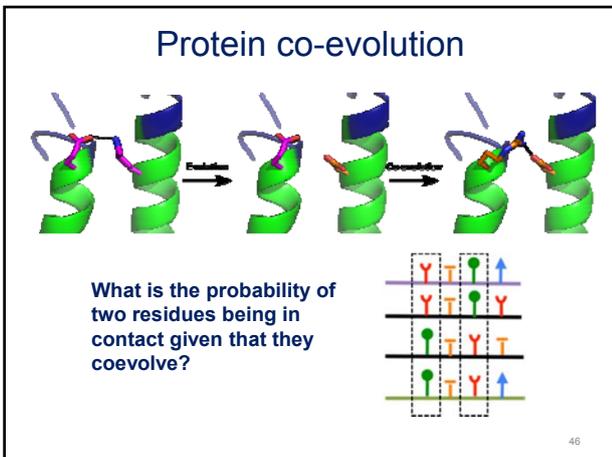
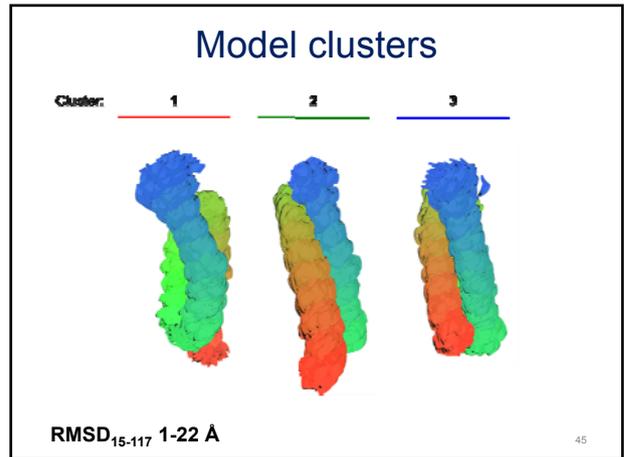
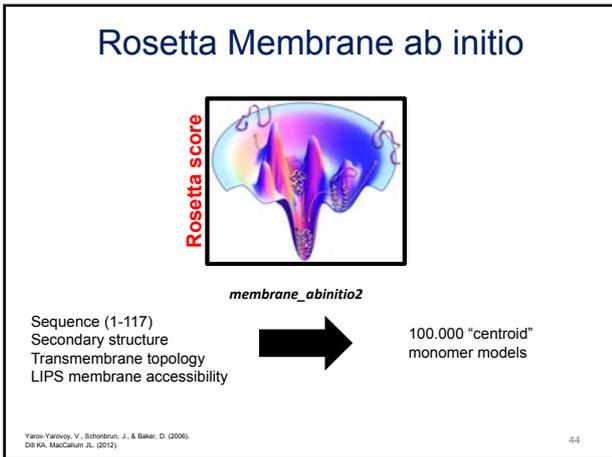
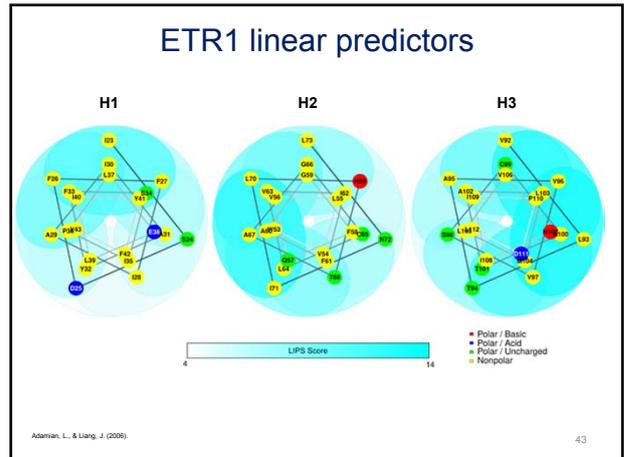
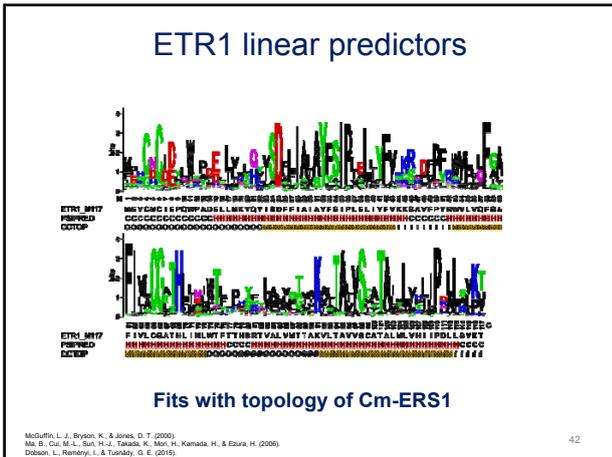


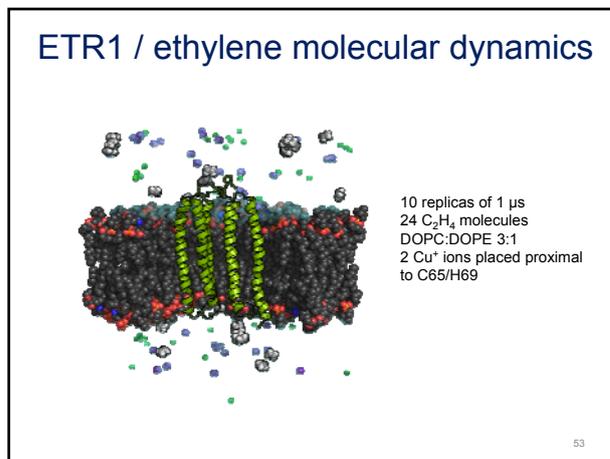
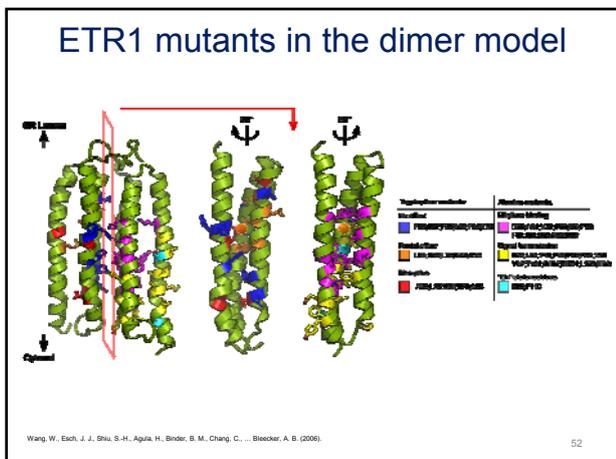
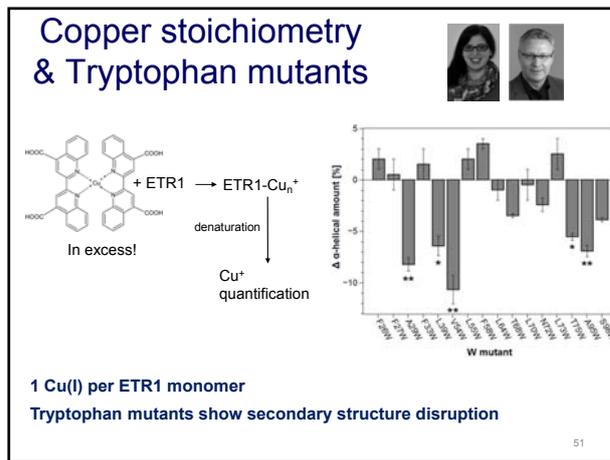
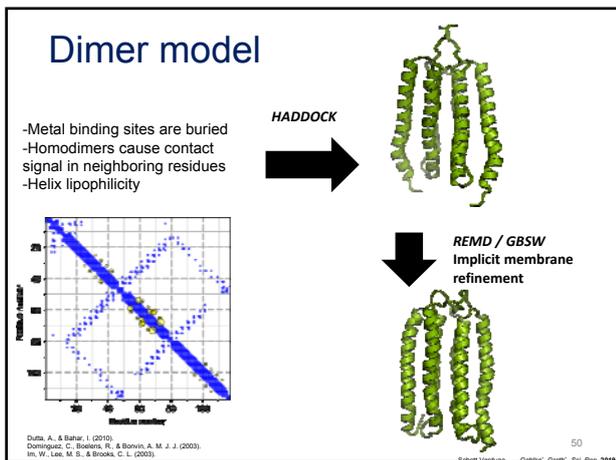
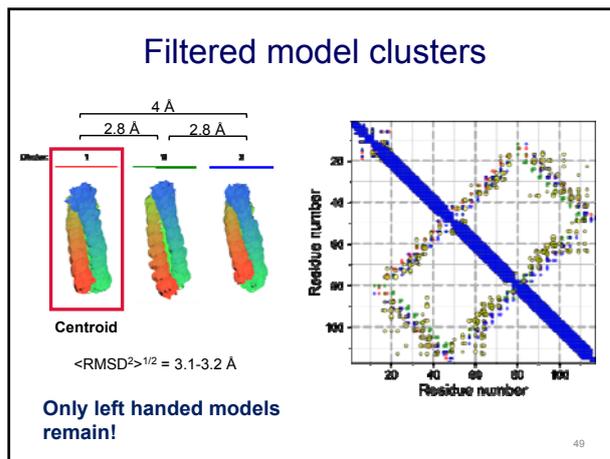
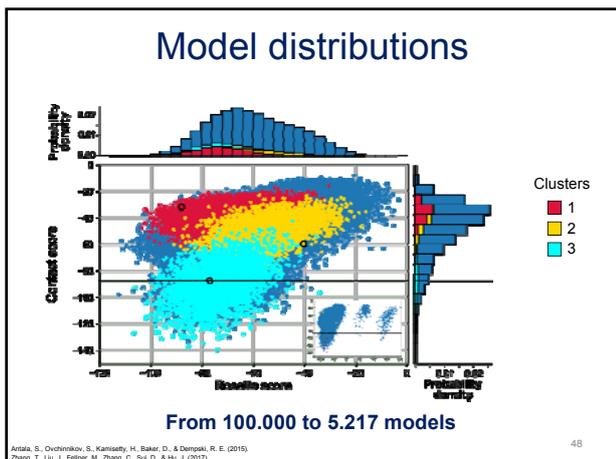
No structure/homolog available!
 Alanine scanning of function related residues*
 288 Cys, 20 Trp, 2 Cys

- 1) Make a structural model of the TM domain (N-ter 117 residues)
- 2) Assemble a dimeric model
- 3) Measure copper stoichiometry & tryptophan scanning (AG Groth)
- 4) Evaluate ethylene binding to the model

Two component system
 ERS1 crystal
 Crystallized
 Crystallized

Mueller-Dieckmann, H. J., Gantz, A. A., & Kim, S. H. (1995).
Wang, W., Esh, J. J., Shu, S.-H., Agila, H., Binder, B. M., Chang, C., ... Becker, A. B. (2006).
Mayerhofer, H., Panesarvelam, S., Kallonen, H., Tuokkanen, A., Mertens, H. D. T., & Mueller-Dieckmann, J. (2016).
Milo, D., Dick, M., Muhias, D., Pfeifer, C., Kinnari, A., Gohlke, H., & Groth, G. (2018).





ETR1 / ethylene binding

Without incorporating ethylene-copper electron exchange!

54

EIN2 - Mechanism of action

TO THE NUCLEUS STABILIZES EBF1/2

57

There are still many unknowns about structural organization of EIN2, but ...

Some of the confirmed phosphorylation sites

Protease cleavage site?

Nuclear localization signal (NLS)

- necessary for EIN2 C-END nuclear localization
- crucial for translational repression of EBF1/2
- required for formation of EIN2-ETR1 complex

58

NLS-based peptides inhibit EIN2-ETR1 complex formation

NLS-based octapeptide:
NOP-1 = H₂N-LKRYKRRL-CONH₂
 Random octapeptide:
ROP-1 = H₂N-EFLYMSVN-CONH₂

59

GAF domain of AtETR1 is involved in binding of the inhibitory peptide NOP-1

Microscale thermophoresis - AtETR1 constructs labeled with thiol-reactive Alexa Fluor 488

Truncated AtETR1 (amino acid ranges)	AtETR1 (subdomains)	Peptide	K _d /nM
full length: 1-738	TM-GAF-DHb-CA-RD	NOP-1	88 ± 41
1-589	TM-GAF-DHb-CA	NOP-1	101 ± 37
1-407	TM-GAF-DHb	NOP-1	83 ± 25
1-307	TM-GAF	NOP-1	104 ± 24
1-157	TM	NOP-1	No binding
306-738	DHb-CA-RD	NOP-1	No binding

62

Model of the GAF domain

Monomer generation

Generated by TopModel
 Evaluated by TopScore

GAF (71% correct)
 Templates
 Flexible regions (~50% error)

66

Model of the GAF domain

Dimer generation

ETR1 forms a dimer mediated by their GAF domains

TopDock (unpublished)

- Struct.-based hom.search
- Predict interface contacts
- Restrained docking

67

Binding site prediction

MD simulations of free ligand diffusion

Three sites predicted

68

Site I is NOT involved in the binding of NOP-1

Predicted NOP-1 binding sites in the GAF domain

Microscale thermophoresis
AtETR1¹⁻³⁰⁷ Ala mutants + NOP-1

Mutant I: $K_d = 128 \pm 65$ nM

- Mutant I (E177A, E178A, E204A, D283A)
- Mutant II (E178A, E204A)
- Mutant III (E178A, E204A)

69

Site III is the binding site of NOP-1

Predicted NOP-1 binding sites in the GAF domain

Tryptophan fluorescence quenching
AtETR1¹⁻³⁰⁷ Trp mutants + NOP-1 (in 10-fold excess)

Sites I II III

Tryptophan fluorescence reporters

- W288F, W288R
- L244W, W265F, W288F, W288R
- Y205W, W265F, W288F, W288R
- T161W, W265F, W288F, W288R
- M148W, W265F, W288F, W288R

70

Potential influence of NOP-1 on ETR1 signal transmission

Structural frameworks

Constraint Network Analysis (CNA)

Proteins as networks

Maxwell 1864; Laman 1970; Tay & Whiteley 1984; Katoh & Tanigawa 2009. Jacobs & Thorpe, Phys. Rev. Lett., 1995 Jacobs et al., Proteins, 2001

71

Potential influence of NOP-1 on ETR1 signal transmission

Clustering of NOP-1 binding modes at site III

73

Potential influence of NOP-1 on ETR1 signal transmission

Hypothesis:
Dimer stability is strengthened
⇒ Signal transmission is attenuated

74 Milić, D., Dick, M., Mulhaes, D., Pflieger, C., Kinnen, A., Gohlke, H., Groth, G., *Sci. Rep.* 2018

Conclusions

- Atomistic model for ETR1 TM sensor domain
- Model shows agreement with experimental mutations
- Predicted ethylene binding pathway
- Identified the interacting domain of NOP-1 – GAF
- Provided a model for how NOP-1 can interfere with signal transmission in ETR1

Outlook

- TM/cytoplasmic domains connection
- Receptor-copper-ethylene interaction
- Full receptor model of signal transmission

76

The global impact of dementia

Alzheimer's disease is the leading cause of dementia

Alzheimer's Association®. 2019 Alzheimer's disease facts and figures, 2019.
Alzheimer's Disease International. World Alzheimer Report 2018, 2018.

The global impact of dementia

Alzheimer's disease is the leading cause of dementia

- Currently, there is no cure for AD
- Changes in the brain begin 20 years or more before any AD-related symptoms are expected to occur
- Key to slowing or stopping the progression of AD is a therapeutic intervention in the very early stages

⇒ It is essential to conclusively recognize AD-related pathological hallmarks

Saravali-Castro, A., et al. *Ann Neurol*, 2017
Bakke, S.J., Harris, J. *EMBO Mol Med*, 2016

Detection of amyloid fibrils

THT and CR are well-known as potent fluorescent dyes for amyloids

The exact nature of how both probes bind to the Aβ(1-42) fibril remained elusive.

Essential for the systematic search for novel molecular probes.

Detection of amyloid fibrils

THT and CR are well-known as potent fluorescent dyes for amyloids

Gremer, L., et al. *Science*, 2017

MD simulations of dye binding

The dyes spontaneously recognize energetically preferred binding epitopes

Aβ(1-42) fibril (at pH 2)[†]

THT

+ ions, water

† The fibrils are grown at pH 2. Gremer, L. et al., Science, 2017

Simulations of THT binding

THT repeatedly binds to and unbinds from the Aβ(1-42) fibril

45 independent MD simulations of 1 μs length each

MD simulations of dye binding

Towards a binding mode model

225,000 conformations from MD ensemble

?

binding mode model structural and energetic characterization

0.0 Time [μs] 1.0

MD simulations of dye binding

Towards a binding mode model

Simulation run

Ligand diffusion

0.0 Time [μs] 1.0

MD simulations of dye binding

Towards a binding mode model

225,000 conformations from MD ensemble

stably bound conformations

binding mode model structural and energetic characterization

THT predominantly binds to the 18VFE22 motif

THT likely binds to the fibril surface

Free energy of binding $\Delta G_{\text{bind.}} = RT \ln(K_D)$ Dissociation constant

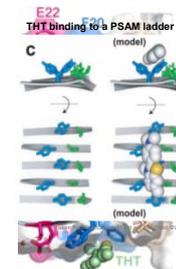
R, T = constants

$K_{D,\text{comp.}}$	$K_{D,\text{exp}}$
112 - 470 nM ($\Delta G_{\text{bind.}} = -9.06 \pm 0.42 \text{ kcal mol}^{-1}$)	790 - 1740 nM (THT to Aβ(1-40) fibrils)

Rodriguez-Rodriguez, C., et al. Metabolism, 2015
Hess, B. L. et al. ChemicalScience, 2009

THT predominantly binds to the 18VFE22 motif

The proposed binding site agrees with experimental observables



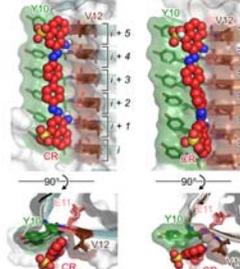
E22
THT binding to a PSAM ladder (model)

C

- Relative fast binding kinetics of THT binding to fibrils suggests that THT has easy accessibility to its binding site. Luikovits, R., et al., Arch Biochem Biophys, 2008
- THT does not bind to monomers; a minimal THT binding site on the fibril surface covers four consecutive β -strands. Bencalones, M., et al., J Mol Biol, 2010
- Similar side chain orientation found for THT binding to peptide self-assembly mimics (PSAM). Bencalones, M., et al., J Mol Biol, 2010
- In amyloid fibrils, THT is suggested to bind to grooves formed by side chains on β -sheets on surface. Podda, M.F.H., et al., J Struct Biol, 2010

CR predominantly binds to the 10YEV12 motif

CR likely binds to the fibril surface



Y10 **M12** **V15** **V12**

+5
+4
+3
+2
+1

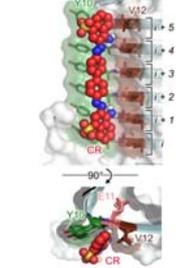
90°

$K_{D,comp.}$	$K_{D,exp}$
0.8 - 70 nM <small>($\Delta G_{bind}^0 = -11.1 \pm 1.31$ kcal mol⁻¹)</small>	48 - 1500 nM <small>(CR to Aβ(1-40) fibrils)</small>
3 - 1458 nM <small>($\Delta G_{bind}^0 = -9.8 \pm 1.8$ kcal mol⁻¹)</small>	

Zhou, Z., et al., J Mol Chem, 2009
Wang, Y., et al., J Labelled Compd Radiopharm, 2010

CR predominantly binds to the 10YEV12 motif

The proposed binding site agrees with experimental observables



Y10 **M12** **V15** **V12**

+5
+4
+3
+2
+1

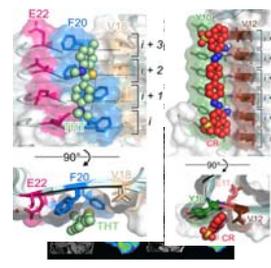
90°

- Different binding sites for THT-like and CR-like ligands. Wessberg, M., J Chem Biol, 2010
- Most studies suggest that the long axis of CR is perpendicular to the direction of the β -strands. Wessberg, M., J Chem Biol, 2010
- The affinity of CR-like probes decreases significantly after replacing the amino by hydroxyl groups. Li, L., Car Med Chem, 2007
- Planarity around the diazo group is likely important. Li, L., Car Med Chem, 2007
- Speculation of multiple CR binding sites
 - complexed by mainly hydrophobic contacts
 - complexed by mainly ionic contactsWessberg, M., J Chem Biol, 2010

Conclusion

Towards improved diagnostic strategies of Alzheimer's disease

- Binding modes for THT and CR to the A β (1-42) fibril were identified by unbiased MD simulations and calculations of binding affinities
- Proposed binding modes agree with previous experimental observations
- The binding modes may provide a starting point for the systematic search and design of novel and improved molecules that bind to A β (1-42) fibrils
- Essential for conclusively diagnosing amyloid fibrils-related diseases *in vitro*



E22 **F20** **V15** **V12**

+5
+4
+3
+2
+1

90°

Acknowledgment

- Daniel Mulnaes
- Stephan Schott-Verdugo
- Lena Müller
- Markus Dick
- Christopher Pfleger
- Dalibor Millic
- Georg Groth

- Benedikt Frieg
- Lothar Gremer
- Henrike Heise
- Dieter Willbold

- Computing time
- Funding



